



## Review Article

## Ethnopharmacological review of ginger for anticancer activity

Amit Gajanan Nerkar<sup>1,2,3,\*</sup>, Srushti Ghadge<sup>1</sup><sup>1</sup>Dept. of Pharmacy, CAYMET's Siddhant College of Pharmacy, Sudumbare, Pune, Maharashtra, India<sup>2</sup>Founder and Director, Ateos Foundation of Science Education and Research, Pune, Maharashtra, India<sup>3</sup>Founder and Director, Carolene Therapeutics, Pvt. Ltd, Aurangabad, Maharashtra, India

## ARTICLE INFO

## Article history:

Received 24-10-2022

Accepted 16-11-2022

Available online 21-11-2022

## Keywords:

Ginger

Ethnopharmacology

Anticancer activity

Pharmacognosy

Zingiber Officinale

## ABSTRACT

Zingiber officinale is a plant found locally in India that has been widely used as a flavouring agent in savoury dishes such as curries and sweets such as cakes and cookies, alcoholic beverages as well as in alcoholic beverages. like in tea. Ginger is a well-known herb, commonly used in traditional medicine all over the world. Ginger has been used for thousands of years to treat colds, nausea, arthritis, migraines, and high blood pressure. The many pharmacological activities of ginger are antiemetic, antidiabetic, analgesic, anti-inflammatory, anticancer, antioxidant, anticoagulant, antibacterial, anti-inflammatory, estrogenic and cardiovascular activities. Chemical irritants and an unsaturated phenolic ketone liquid C17H24O3 are responsible for the spicy taste of ginger. The main components of ginger are aromatic essential oils, antioxidants and pungent resins. These aromatic or pungent compounds have been identified as C6H5C(O)CH3, known as a chemical irritant, liquid unsaturated phenolic ketones C17H24O3 and Vanillylacetone.

This is an Open Access (OA) journal, and articles are distributed under the terms of the [Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License](https://creativecommons.org/licenses/by-nc-sa/4.0/), which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: [reprint@ipinnovative.com](mailto:reprint@ipinnovative.com)

## 1. Introduction

Ginger (Canton ginger) belongs to the family Zingiberacea.<sup>1</sup> It is herbaceous perennial plant.<sup>2</sup> It is commonly used as a spice and medicinal plant.<sup>2</sup> The part of the plant used is the rhizome. The plant produces an orchid-like flower with purple-flecked yellow-green petals. Ginger (*Zingiber officinale* (L.) Roscoe) has been used as a spice for over 2000 years.<sup>3</sup> Ginger contains up to 3% of the essential oils responsible for the aroma of the spice.<sup>4</sup> In India and countries with hot and humid climates, ginger is consumed in large quantities and is good for digestive problems.<sup>5</sup> Ginger belongs to the plant family that includes cardamom and turmeric. Its spicy taste is mainly due to the presence of ketones, especially ginger root, which seems to be the main component of ginger studied in most of

the health-related scientific studies.<sup>6</sup> The rhizome, which is the horizontal stem from which the roots grow, is the main part of the ginger that is consumed. Ginger's current name comes from the Middle English *gingivere*, but the spice dates back more than 3,000 years to the Sanskrit word *srngaveram*, meaning "horny root", based on its shape.<sup>7</sup> In Greek it is called *ziggiberis*, and in Latin *zinziberi*. Interestingly, ginger does not grow in the wild and its true origin is uncertain. The Indians and Chinese are said to have produced ginger as a root tonic for over 5000 years to cure many ailments, and the herb is now grown throughout the humid tropics. India is the largest ginger producer. Ginger was used as a salting agent long before official history was recorded. It was an extremely important trade item and was exported from India to the Roman Empire over 2000 years ago, where it was particularly prized for its healing properties. Ginger continued to be a much

\* Corresponding author.

E-mail address: [dragnerkar@gmail.com](mailto:dragnerkar@gmail.com) (A. G. Nerkar).

sought-after commodity in Europe even after the fall of the Roman Empire, with Arab traders controlling the trade in ginger and other spices for centuries.<sup>8</sup> In the 13th and 14th centuries, the value of a pound of ginger was equivalent to the price of a sheep. Raw and preserved ginger was brought to Europe in the Middle Ages, where it was included in the official accumulations of various countries. During the Middle Ages, it was imported in canned form for use in sweets. Queen Elizabeth I of England is credited with inventing the gingerbread man, which has become a popular Christmas treat.<sup>9</sup>

### 1.1. Classification

Taxonomic	Dry ginger plant
Domain	Eukaryota
Kingdom	Plantae
Phylum	Spermatophyta
Subphylum	Angiosperms
Class	Monocotyledon
Order	Zingiberales
Family	Zingiberaceae
Genus	Ginger
Species	Zingiber officinale

### 1.2. Active Constituents

At least 115 components of fresh and dried ginger varieties have been identified through multiple analytical procedures. Ginger has many active components, such as phenolics and terpenoids.<sup>10</sup> The phenolic compounds in ginger are mainly ginger root, <sup>6</sup>-shogaol and paradols. In fresh ginger, ginger root has the main polyphenols such as 6-gingerol, 8-gingerol and 10-gingerol. Ginger root is the major constituent of fresh ginger and was found to be slightly reduced in dried ginger, while concentrations of <sup>6</sup>-shogaol, which is the main dehydration product of gingerol, were more abundant<sup>11</sup> in drier ginger than fresh ginger. At least 31 compounds related to gingerol have been identified from crude methanolic extracts of fresh ginger rhizomes (Jiang, Solyom et al. 2005). Ginger is rich in active ingredients, such as phenolics and saponins. Ginger has been subdivided into at least 14 bioactive compounds, including z<sup>4</sup>-gingerol, <sup>6</sup>-gingerol, <sup>8</sup>-gingerol, <sup>10</sup>-gingerol, <sup>6</sup>-paradol, <sup>12</sup>-shogaol, <sup>6</sup>-shogaol, 1-dehydro-<sup>10</sup>-gingerdione, <sup>10</sup>-gingerdione, 3-heptanone, 5-hydroxy-1,7-bis (4-hydroxy-3-methoxyphenyl), C<sub>21</sub>H<sub>24</sub>O<sub>6</sub>, linear log heptanoids, 1, 7-bis- (4' hydroxyl-3' methoxyphenyl)-5-methoxyheptan-3-one and methoxy-<sup>10</sup>-gingerol.<sup>13</sup> The proportions of each individual ingredient in a sample of ginger depend on the country of origin, commercial processor and fresh, dried or processed ginger.<sup>14</sup> Among the pungent bioactive

components of Jamaican ginger, including, <sup>6</sup>-, <sup>8</sup>-, and <sup>10</sup> roots and <sup>6</sup>-shogaol, <sup>6</sup>-gingerol appears to be the pungent compound. has the most abundant biological activity in most of the petroleum resins.<sup>12</sup> Although phylogenetic analysis revealed that all ginger samples from different geographical origins were genetically indistinguishable, metabolic profiling revealed quantitative differences. on the content of <sup>6</sup>-, <sup>8</sup>- and <sup>10</sup>- ginger root.<sup>15</sup> An evaluation of the concentrations of <sup>6</sup>-, <sup>8</sup>-, and <sup>10</sup> root and <sup>6</sup>-shogaol in 10 different ginger root dietary supplements randomly purchased from pharmacies. Various drug and grocery stores, natural foods have given amazing results. Perhaps unsurprisingly, the amounts of these active ingredients vary widely, from none or very small amounts to several milligrams per gram. In addition, the recommended dietary intake ranges from about 250 mg to 4.8 g/day. The basis for the wide dosage range is unclear. These studies show that ginger contains many bioactive compounds and standardization of the content is very lacking.<sup>16</sup>

## 2. Ethnopharmacological review of anticancer activity

Vallinoids in Ginger possess anticancer activities. Vallinoids are certain pungent active constituents like <sup>6</sup>-gingerol and <sup>6</sup>-paradol, and some other constituents like <sup>6</sup>-shogaol, vanillylacetone etc. It was demonstrated that <sup>6</sup>-gingerol, <sup>6</sup>-shogaol and vanillylacetone possess anti-angiogenesis dependent human diseases properties that helps to fight the cancer. There has been substantial research on the anticancer activities of active constituents of ginger and its various components and were reviewed the properties of ginger and its chemoprophylaxis effects of numerous dietary and medicinal plants. Studies were conducted on anticancer activities of various parts and forms of ginger especially <sup>6</sup>-gingerol, <sup>6</sup>-shogaol, especially <sup>6</sup>-shogaol; and zerumbone, a sesquiglucoside compound derived from ginger and a number of minor components and metabolites.<sup>17-21</sup> The effectiveness of ginger in preventing or suppressing cancer growth has been examined in a variety of cancer types such as lymphoma, hepatoma, colorectal cancer, breast cancer, skin cancer, liver cancer, and bladder cancer. The mechanisms showed the anticancer activities of ginger and its components include antioxidant activity and induction of apoptosis, decrease proliferation, cause cell-cycle arrest, and suppress activator protein 1 (AP-1) and NF- $\kappa$ B/COX-2 indicating pathways.<sup>22</sup>

The antitumor activities of <sup>6</sup>-gingerol and zerumbone are related to their antioxidant activities. Several components of ginger were reported to have potent anticancer promoter activity based on their ability to inhibit TPA-induced Epstein-Barr virus early antigen (EBV-EA) in B lymphocytes in humans.<sup>6,23</sup> -Gingerol was reported to block the potent invasive capacity of reactive oxygen species of AH109A liver cancer cells ascites by reducing peroxide levels.<sup>24</sup> In normal mouse RL34 hepatic epithelial

**Table 1:** It shows anticancer activity of ginger with mechanism of action

Compound Name	Cancer	Mechanism	Cell Lines/System
Ginger extract	Liver cancer	Reduced the elevated expression of TNF- $\alpha$ and NF-kB and tumor growth	In-vitro
Ginger-whole and <sup>6</sup> -gingerol	Non-small-cell lung cancer cells	Release of cytochrome c	In-vitro
$\beta$ -Elemence	Non-small-cell lung cancer cells	Release of cytochrome c	In-vitro
6-gingerol	Breast cancer	Inhibites cell adhesion invasion motility	In-vitro
	Skin cancer	Enhance apoptosis	Mouse
	Colon cancer	Inhibition of leukotriene activity	Mice
	Lung and colon cancer	Suppresses modulatory mechanism of growth and induces apoptosis, Reduces expression of NF-kB	Mouse
Zerumbone	Colon cancer	Activation of extraction signal-regulated kinase $\frac{1}{2}$ p38 mitogen-activated protein kinase	In-vitro
	Osteoclastogenesis	Blocks NF-kappa B expression.	Mouse nonocyte
6-Shogaol	Lungs cancer	Inhibition of AKT	In-vitro
6-Shogaol	Breast cancer	Anti-metastasis	In-vitro
Enone-diaryl heptanoid, 6-shogaol, <sup>10</sup> -gingerol	Liver/against nine human tumor cell (lines)	Inhibition of lipid peroxidation Antioxidant activity, cytotoxic	In-vitro
Terpenoids	Endometrial Cancer Cell	Induces apoptosis by activation of p53	In-vitro
6-Shogaol	Cancer cell	Anticancer	In-vitro

cells, zerumbone was found to induce glutathione S-transferase and localize the transcription factor Nrf2, which binds to the antioxidant response factor gene (ARE) of the phase II enzyme.<sup>25</sup>

Zerumbone promotes the expression of several Nrf2/ARE-dependent phase II enzyme genes, including  $\gamma$ -glutamyl-cysteine synthetase, glutathione peroxidase, and hemeoxygenase-1.<sup>26</sup> Others have reported that zerumbone reduces TPA-induced hydrogen peroxide formation and water retention in proportion to increased levels of various antioxidant enzymes.<sup>26</sup> These types of alterations were associated with decreased incidence of 7,12-dimethylbenz[a]anthracene (DMBA)-inducing/promoting TPA tumors, number of tumors per mouse, and tumor volume.<sup>27</sup>

Zerumbone has also been reported to downregulate CXCR4 chemokine receptor 4 (CXCR4), which is abundantly expressed in various tumors including breast, ovarian, prostate, gastrointestinal tract, head and neck, bladder optic nerve, brain and melanoma.<sup>28</sup> Because CXCR4 mediates the migration of tumor cells to specific organs expressing its ligand, CXCL12, zerumbone has been suggested as a potent and potent cancer metastasis inhibitor effective in blocking CXCR4 in a variety of cancers, including pancreatic, lung, kidney, and skin cancers.<sup>28</sup> In addition, zerumbone effectively attenuated human mammary gland tumor cell cytotoxicity and multiple myeloma-induced osteoclastogenesis and dose-dependently reduced bone resorption in MDA-bearing Athiorhodaceae nude mice -

MB-231, suggests that it may be effective in preventing bone cancer-related. bone loss or osteoporosis.<sup>6,29</sup> Gingerol has also been reported to suppress adhesion, invasion, mobility, matrix metalloproteinase (MMP)-2 and MMP-9 messenger ribonucleic acid (mRNA) expression and protein activities. in the human breast cancer cell line MDA-MB-231.<sup>30,31</sup>

Ginger and its components have been reported to inhibit tumor promotion in rat skin.<sup>32</sup> In particular, <sup>6</sup>-gingerol was found to be highly effective as an anti-skin cancer agent in vivo in a rat skin model encouraging two-step initiation. In this model, tumors were initiated with a single application of DMBA, followed by repeated topical TPA applications beginning several days later.<sup>33</sup> Topical application of <sup>6</sup>-gingerol on the shaved backs of female ICR rats reduces the rate of DMBA-initiated/TPA-promoted skin papilloma formation and also inhibits epidermal ornithine decarboxylase activity and inflammation caused by TPA.<sup>34</sup> The results of a similar study indicated that in the DMBA/TPA skin tumor model, the application of <sup>6</sup>-paradol or <sup>6</sup>-dehydroparadol before TPA application significantly reduced the number of tumors per mouse. and the number of mice with tumors.<sup>35</sup>

Previous studies showed that gingerol was an effective inhibitor of azoxymethane-induced intestinal carcinogenesis in rats.<sup>36</sup> Ginger supplementation (50 mg/kg body weight) has been reported to reduce the number of tumors as well as the incidence of 1,2-dimethylhydrazine (DMH)-induced colon cancer.<sup>37</sup> The effect is attributed to decreased oxidative damage associated with increased

activity of catalase, superoxide dismutase, glutathione peroxidase and glutathione transferase as well as increased GSH.<sup>37</sup> This group then reported that administering ginger to mice treated with DMH significantly reduced the incidence and number of tumors as well as the activity of microbial enzymes (GUSB) and mucopolysaccharidase.<sup>37</sup> Finally, Wistar rats fed ginger extract (1% dietary blend) showed a significantly lower number of ureteral lesions (hyperplasia and neoplasia) compared with the untreated groups.<sup>38</sup>

Studies show that compounds in ginger prevent the proliferation of human cancer cells through the induction of apoptosis.<sup>39,40</sup> A salt extract prepared from ginger extract suppressed HEP-2 cell proliferation by inducing cytotoxic effects and DNA fragmentation.<sup>41</sup> Ginger extract and especially<sup>6</sup> -gingerol were reported to effectively reduce YYT colonC.<sup>10,42</sup> -Gingerol has been reported to induce a significant and prolonged increase in intracellular calcium and cytotoxicity in human colorectal cancer SW480 cells.<sup>6,43</sup> -Gingerol has been reported to inhibit both proliferation and infiltration of AH109A ascites cells and appears to work by inducing S phase arrest, prolonging cell replication time. of liver tumors and increased apoptosis.<sup>44</sup> This compound also induces cell cycle arrest and suppresses the growth of human pancreatic cancer cell lines, human pancreatic carcinoma (HPAC) cells, which express wild-type p53 and BxPC-3 cells expressing a mutant p53 protein.<sup>45</sup> Interestingly,<sup>6</sup> gingerol appeared to be most effective in inducing apoptosis in p53 mutant cells and inducing arrest, but not apoptosis in epithelial cells. present p53.<sup>6</sup> -Gingerol was further reported to suppress proliferation and induce apoptosis or G1 cell cycle arrest in several colorectal cell lines, including HCT116, SW480, HT29 cells, LoVo and Caco2.<sup>46</sup> These effects are associated with decreased levels of cyclin D1 (a proto-oncoprotein overexpressed in cancer) and increased expression of the nonsteroidal anti-inflammatory drug (NSAID) activating gene (NAG-1), a protein autophagy and antibiotics.<sup>46</sup>

Through the comparison of promotion-sensitive (P+) and promotion-resistant (P-) derivatives of the JB6 mouse epidermal cell lines, AP-1 was reported to play an important role in promoting promote tumorigenesis.<sup>47</sup> In addition, blocking promoter-induced AP-1 activation inhibited neoplastic transformation<sup>48</sup> Epidermal growth factor (EGF) is known to induce AP-1 activity and cell transformation at relatively high levels.<sup>47</sup> Previously, we investigated the effects of two structurally related compounds from the ginger family,<sup>6</sup> -gingerol and<sup>6</sup> -paradol, on EGF-induced cellular transformation and activation. AP.1 chemistry . Our results provided the first evidence that both compounds block EGF-induced cellular transformation, but through different mechanisms.<sup>6</sup> -Gingerol appears to act by directly inhibiting AP-1 DNA-binding activity and metabolism, while<sup>6</sup> -

paradol appears to act by inducing apoptosis<sup>49,50</sup> Others reported that<sup>6</sup> -gingerol induced DNA fragmentation and suppressed Bcl-2 expression in HL-60 myeloid leukemia cells (Wang et al. 2003), and also induced growth inhibition and caspase-mediated apoptosis in human squamous cell carcinoma A431 cells.<sup>6,51,52</sup> -paradol and other structurally related derivatives, such as<sup>10</sup> -paradol,<sup>3</sup> -dehydroparadol,<sup>6</sup> -dehydroparadol and<sup>10</sup> -dehydroparadol, inhibited proliferation cells of KB oral squamous cell carcinoma in record time. and is dose dependent.<sup>6,53</sup> -dehydroparadol was more potent than the other compounds tested and induced apoptosis by a caspase-3-dependent mechanism.<sup>6,52</sup> -Shogaol [1-(4-hydroxy-3-methoxyphenyl)-4-lie-3-one], a ketone from ginger, exhibits the strongest cytotoxicity against humans A549, SK-OV -3, SK-MEL-2, and HCT15 tumor cells, compared with<sup>4</sup> -<sup>6</sup> -,<sup>8</sup> -, and<sup>10</sup> -gingerroot.<sup>54</sup> This compound also inhibited the proliferation of several transgenic mouse ovarian cancer cell lines, including C1 and C2.<sup>54</sup> In addition,<sup>6</sup> -shogaol has been reported to inhibit growth and induce apoptosis in COLO 205 cells.<sup>55</sup> Treatment with<sup>6</sup> -shogaol, but not<sup>6</sup> -gingerol, induces DNA fragmentation in COLO 205 colon cancer cells. Death is mediated by activation of caspase-9, -3 and -8, leading to mitochondrial cytochrome c release, modulation of pro-apoptotic Bax and negative effects of anti-apoptotic Bcl2, and induction of growth arrest and DNA damage (GADD)-factor transcribed 153 (GADD153) mRNA and protein.<sup>55</sup>

NF- $\kappa$ B is a rapidly induced stress-responsive transcription factor that enhances the transcription of many genes, including cytokines, growth factors, and acute-responsive proteins.<sup>56</sup> Its activation is also implicated in the mitogen-activated protein (MAP) kinase signaling pathways.<sup>57</sup> The mechanism of NF- $\kappa$ B activation is well known. In its inactive form, NF- $\kappa$ B found in the cytosol binds to an inhibitory protein known as kappa inhibitor B (I $\kappa$ B). When stimulated, I $\kappa$ B is phosphorylated by an I $\kappa$ B kinase, releasing it from NF- $\kappa$ B and subsequently degraded. After detaching from I $\kappa$ B, NF- $\kappa$ B is transferred to the nuclear region, where it activates gene transcription by binding to its specific DNA sequence present in certain genes. Importantly, NF-B activation has been implicated in the initiation or acceleration of tumorigenesis,<sup>58</sup> and in JB6 cells, NF- $\kappa$ B inhibition also suppresses tumorigenesis. cell changes induced by tumor promoters (60, Li et al. 1997).<sup>6</sup> -Gingerol may exert its effects by blocking the NF- $\kappa$ B/COX-2 pathway. This idea is supported by data indicating that the reduction of UVB-induced expression and COX-2 metabolism by<sup>6</sup> -gingerol is associated with inhibition of I $\kappa$ B $\alpha$  (Ser32) phosphorylation leading to to reduce NF- $\kappa$ B translocation from the cytosol to the nucleus in HaCaT cells.<sup>59</sup> A ginger extract given to mice with experimentally induced liver cancer reduced NF- $\kappa$ B and TNF- $\alpha$ .<sup>6,60</sup> -Gingerol has been reported to block

TNF-linked apoptosis.

Zerumbone has been reported to block NF- $\kappa$ B activation induced by a variety of stimuli, including tumor necrosis factor (TNF), cigarette smoke condensation, and hydrogen peroxide.<sup>61</sup> It also blocks phosphorylation and degradation of I $\kappa$ B $\alpha$  kinase, leading to downregulation of constitutively active NF- $\kappa$ B and some of its upregulated gene targets, such as COX-2, cyclin D1, Bcl2. and anti-apoptotic genes, thereby enhancing chemotherapy-induced cell death.<sup>62</sup> Zerumbone has also been reported to block the activity of the NF- $\kappa$ B ligand receptor activator (RANKL) in mouse monocytes (progenitor cells of osteoclasts) by inhibiting the activity, phosphorylation and degradation of kinase I $\kappa$ B $\alpha$  (Sung et al. 2009). Oral administration of zerumbone (100, 250, or 500 ppm) to ICR mice reduces the inflammation and diversity of colon adenocarcinoma induced by intraperitoneal injection of azoxymethane (AOM, 10 mg/kg body weight; Kim et al. 2009). In addition, zerumbone (250 or 500 ppm) effectively suppressed 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone-induced lung adenoma formation in female A/J mice.<sup>52</sup> This ginger derivative appears to exert its effects through inhibiting proliferation, inducing apoptosis, and inhibiting the expression of NF-B and heme oxygenase in colon cancer tissues. and lungs<sup>63</sup> In a previous study,<sup>6</sup> -gingerol was reported to be able to inhibit both vascular endothelial growth factor (VEGF) and basic fibroblast growth factor (bFGF) inducing proliferation. human endothelial cells and induce cell cycle arrest in the G1 phase.<sup>6,64</sup> -Gingerol also suppresses capillary-like tubule formation by endothelial cells in response to vascular endothelial growth factor (VEGF), and strongly inhibits endothelial cell germination. in the rat aorta and the formation of new blood vessels in the rat cornea in response to VEGF.<sup>65</sup>

Researchers have suggested that the effectiveness of ginger may be related to its ability to inhibit prostaglandin and leukotriene biosynthesis.<sup>66</sup> Some researchers have shown that gingerol actively inhibits arachidonate 5-lipoxygenase, an enzyme in leukotriene biosynthesis.<sup>67</sup> The protein leukotriene A4 hydrolase (LTA4H) was considered a suitable target for cancer therapy and our prediction in silico using reverse splicing suggested that LTA4H could be a potential target for<sup>6</sup> - gingerol.<sup>68</sup> The prediction is supported by the study that<sup>6</sup> -gingerol suppresses the growth of cancer cells independent of the anchorage by binding to LTA4H and inhibiting the activity of LTA4H in HCT116 colorectal cancer cells. It was also found that<sup>6</sup> -gingerol effectively suppressed tumor growth in vivo in nude mice, an effect mediated by inhibition of LTA4H activity. Taken together, these results point to an important role for LTA4H in cancer and also support the antitumor efficacy of<sup>6</sup> -gingerol targeting LTA4H in preventing colorectal cancer.<sup>68</sup> More importantly, these are the first results to identify a direct target of<sup>6</sup> -gingerol to explain its antitumor activity.<sup>69</sup>

It was found that when cultured ovarian cancer cells were treated with<sup>6</sup> -shogaol, they exhibited marked inhibition associated with NF- $\kappa$ B inhibition as well as inhibition angiogenic factors, VEGF and IL-8. Therefore, this compound plays an active role in preventing cancer angiogenesis.<sup>70</sup> Furthermore, it was concluded that people consuming dietary ginger (0.5% or 1.0%) showed no activity on malformation of aberrant crypt foci formation (ACF). or decreased proliferation index or crypt cells of the apoptotic colon induced by DMH. in DMH-treated rats. Furthermore, as demonstrated,<sup>70,71</sup> ginger extract could not inhibit the growth of N-butyl-N-(4-hydroxybutyl)-nitrosamine (BBN)/N-methyl- N-nitrosourea (MNU)-induced bladder cancer in male Swiss rats and in mice treated with BBN/MNU/2% ginger, the incidence of grade 2 translocation cell carcinoma was increased.<sup>72</sup>

### 3. Conclusion

Ginger and its active components inhibit growth and proliferation of many cancer cells. Also 6-gingerol and other active constituents in ginger inhibited the growth of many cell-lines and showed cyto-toxic properties. Thus, the anticancer properties of ginger described in this review may give a new direction for anticancer drug discovery.

### 4. Source of Funding

AGN for funding this project through Ateos Foundation of Science Education and Research, Pune, M.S., India

### 5. Conflict of Interest

None.

### 6. Acknowledgement

The Author SG thanks AGN for funding this project through Ateos Foundation of Science Education and Research, Pune, M.S., India, also waiver for the publications and acknowledges the same.


### References

1. Lim TK, Officinale Z. In Edible medicinal and non-medicinal plants. Springer; 2016. p. 469–560.
2. Vashist H, Gupta A, Beri C, Sharma RB. A Report on Aloe vera and turmeric as herbal medicine and cosmetics. *PHARMACIA* . 2014;2(2):60–74.
3. Vasconcelos DS, Mirele EF, Mota NF, Gomes-Rochette DS, Nunes-Pinheiro S, Nabavi DM. Ginger (zingiber officinale roscoe). Academic Press; 2019. p. 235–9.
4. Nweze EI, Okafor JI, Njoku O. Antimicrobial activities of methanolic extracts of Trema guineensis (Schumm and Thorn) and Morinda lucida benth used in Nigerian. *Bio-research*. 2004;2(1):39–46.
5. Rajathi A, Angeline A, Sundarraj S. Processing and medicinal uses of cardamom and ginger-a review. *J Pharm Sci Res*. 2017;9(11):2117–22.
6. Khan S, Pandotra P, Qazi AK, Lone SA, Muzafar M, Gupta AP. Medicinal and nutritional qualities of zingiber officinale. Academic

- Press; 2016. p. 525–50.
7. Vernin G, Parkanyi C. Chemistry of ginger. CRC Press; 2016. p. 107–200.
  8. Abubacker AT. Export potential of ginger.; 2009. Available from: <https://d1wqtxts1xzle7.cloudfront.net/31378319/Ginger-Apr09-with-cover-page-v2.pdf?>
  9. Bode AM, Dong Z. The amazing and mighty ginger; 2011. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK92775/>.
  10. Beta HB, Qian-Qian XY, Xu SY, Cao RY, Gan H, Corke T. Bioactive Compounds and Bioactivities of Ginger (*Zingiber officinale* Roscoe). *Food*. 2019;8(6):185.
  11. Jolad SD, Lantz RC, Chen GJ, Bates RB, Timmermann BN. Commercially processed dry ginger (*Zingiber officinale*): composition and effects on LPS-stimulated PGE2 production. *Phytochemistry*. 2005;66(13):1614–35.
  12. Bailey-Shaw YA, Lawrence AD, Williams CE, Green S, Rodney AM. In-vitro evaluation of the anti-inflammatory potential of selected Jamaican plant extracts using the bovine serum albumin protein denaturation assay. *Int J Pharm Sci Rev Res*. 2017;47(1):145–53.
  13. Han EJ, Kim JM. Quality characteristics of yanggaeng prepared with different amounts of ginger powder. *J East Asian Soc Dietary Life*. 2011;21(3):360–6.
  14. Schwertner HA, Deborah C, Rios JE. Variation in concentration and labeling of ginger root dietary supplements. *Obstet Gynecol*. 2006;107(6):1337–43.
  15. Jiang H, Xie Z, Koo HJ, Mclaughlin SP, Timmermann BN, Gang DR. Metabolic profiling and phylogenetic analysis of medicinal Zingiber species: Tools for authentication of ginger. *Phytochemistry*. 2006;67(15):1673–85.
  16. Schwertner HA, Deborah C, Rios JE. Variation in concentration and labeling of ginger root dietary supplements. *Obstetrics Gynecol*. 2006;107(6):1337–43.
  17. Surh YJ, Lee E, Lee JM. Chemoprotective properties of some pungent ingredients present in red pepper and ginger. *Mutation Res/Fundamental Mol Mech Mutagenesis*. 1998;402(1-2):259–67.
  18. Surh YJ, Park KK, Chun KS, Lee LJ, Lee E, Lee SS. Anti-tumor promoting activities of selected pungent phenolic substances present in ginger. *J Environ Pathol Toxicol Oncol*. 1999;18(2):131–9.
  19. Bode AM, Dong Z. Ginger. CRC Press; 2004. p. 157–68.
  20. Dwivedi M, Sharma S, Shukla P, Mishra SR, Paliwal ASK. Development and evaluation of anticancer polymeric nanoformulations containing curcumin and natural bioenhancers. *J Biomat Tissue Eng*. 2014;4(3):198–202.
  21. Kunnumakkara AB, Koca C, Dey S, Gehlot P, Yodkeeree S, Danda D. Development and Evaluation of Anticancer Polymeric Nano-Formulations Containing Curcumin and Natural Bioenhancers. *J Biomater Tissue Eng*. 2009;4(3):1–24.
  22. Dwivedi, Monika, Shweta. Protective and therapeutic potential of ginger (*Zingiber officinale*) extract and [6]-gingerol in cancer: A comprehensive review. *Phytotherapy Res*. 2018;32(10):1885–907.
  23. Shukla Y, Singh M. Cancer preventive properties of ginger: a brief review. *Food Chem Toxicol*. 2007;45(5):683–90.
  24. Sharma V, Rao LM. A thought on the biological activities of black tea. *Crit Rev Food Sci Nut*. 2009;49(5):379–404.
  25. Nakamura Y, Yoshida C, Murakami A, Ohigashi H, Osawa T, Uchida K. Zerumbone, a tropical ginger sesquiterpene, activates phase II drug metabolizing enzymes. *FEBS letters*. 2004;572(1-3):245–50.
  26. Yang HL, Lee CL, Korivi M, Liao JW, Rajendran P, Wu JJ. Zerumbone protects human skin keratinocytes against UVA-irradiated damages through Nrf2 induction. *Biochem Pharmacol*. 2018;148:130–46.
  27. Ramadan MM, Effat M, El-Shershaby MF. The protective effect of ginger on ovotoxicity induced by 7, 12-dimethylbenz [a] anthracene (DMBA) in rat. *Egypt J Experi Biol*. 2009;5(0):227–33.
  28. Zainal NS, Gan CP, Lau BF, Yee PS, Tiong KH, Rahman ZA. Zerumbone targets the CXCR4-RhoA and PI3K-mTOR signaling axis to reduce motility and proliferation of oral cancer cells. *Phytomedicine*. 2018;39:33–41.
  29. Anandasadagopan S, Kumar P, Sivaprakasam S, Pathmanapan A, Pandurangan TK. Targeting the Key Signaling Pathways in Breast Cancer Treatment Using Natural Agents. *Biomed Pharmacoter*. 2020;150:137–83.
  30. Kaur P, Indu PK, Kondepudi KK, Bishnoi M. Anticancer potential of ginger: mechanistic and pharmaceutical aspects. *Curr Pharm Des*. 2016;22(27):4160–72.
  31. Ma RH, Ni ZJ, Zhu YY, Thakur K, Zhang F, Zhang YY, et al. A recent update on the multifaceted health benefits associated with ginger and its bioactive components. *Food Funct*. 2021;12(2):519–42.
  32. Zheng Y, Geng S, Meng M, Du Z, Yao J, Zhang Q, et al. The Edible Adlay Seed Oil Containing Cinnamaldehyde and Gingerol Prevents DMBA/TPA-Induced Skin Carcinogenesis by Cooperative Ablation of Lipid Rafts. *J Carcinog Mutagenesis*. 2018;8(4):1–7.
  33. Park KK, Chun KS, Lee JM, Lee SS, Surh YJ. Inhibitory effects of [6]-gingerol, a major pungent principle of ginger, on phorbol ester-induced inflammation, epidermal ornithine decarboxylase activity and skin tumor promotion in ICR mice. *Cancer lett*. 1998;129(2):139–44.
  34. Yusof YAM. Gingerol and Its Role in Chronic Diseases. *Adv Exp Med Biol*. 2016;978(3):177–207.
  35. Keum Y, Sam J, Kim K, Lee KH, Park YJ, Surh JM. Induction of apoptosis and caspase-3 activation by chemopreventive [6]-paradol and structurally related compounds in KB cells. *Cancer lett*. 2002;177(1):41–7.
  36. Yoshimi N, Sugie S, Tanaka T, Aijin W, Bunai Y, Tatematsu A. A rare case of serous cystadenocarcinoma of the pancreas. *Cancer*. 1992;69(10):2449–53.
  37. Manju V, Nalini N. Chemopreventive efficacy of ginger, a naturally occurring anticarcinogen during the initiation, post-initiation stages of 1, 2 dimethylhydrazine-induced colon cancer. *Clin Chim Acta*. 2005;358(1-2):60–7.
  38. Ihlaseh S, Muhammad M, De Oliveira EC, Teràn J, De Camargo LLV. Chemopreventive property of dietary ginger in rat urinary bladder chemical carcinogenesis. *World J Urol*. 2006;24(5):591–6.
  39. Surh YJ, Lee E, Lee JM. Chemoprotective properties of some pungent ingredients present in red pepper and ginger. *Mutat Res*. 1998;402(1-2):259–67.
  40. Lee E, Surh YJ. Induction of apoptosis in HL-60 cells by pungent vanilloids. *Cancer lett*. 1998;134(2):163–8.
  41. Padma V, Viswanadha SD, Christie K, Ramkuma M. Induction of apoptosis by Ginger in HEP-2 cell line is mediated by reactive oxygen species. *Basic Clin Pharmacol Toxicol*. 2007;100(5):302–7.
  42. Baliga M, Shrinath R, Jerome D, Manisha M, Pereira J. Update on the chemopreventive effects of ginger and its phytochemicals. *Crit Rev Food Sci Nutr*. 2011;51(6):499–523.
  43. Chen CY, Li YW, Kuo SY. Effect of [10]-gingerol on [Ca<sup>2+</sup>]<sub>i</sub> and cell death in human colorectal cancer cells. *Molecules*. 2009;14(3):959–69.
  44. Yagihashi S, Miura Y, Yagasaki K. Inhibitory effect of gingerol on the proliferation and invasion of hepatoma cells in culture. *Cytotechnology*. 2008;57(2):129–36.
  45. Pramanik KC, Boreddy SKSR. Role of mitochondrial electron transport chain complexes in capsaicin mediated oxidative stress leading to apoptosis in pancreatic cancer cells. *PloS one*. 2011;6(5):20151.
  46. Lee SH, Cekanova M. Multiple mechanisms are involved in 6- gingerol-induced cell growth arrest and apoptosis in human colorectal cancer cells. *Mol Carcinog*. 2008;47(3):197–208.
  47. Huang C, Ma WY, Bowden GT, Dong Z. Ultraviolet B-induced activated protein-1 activation does not require epidermal growth factor receptor but is blocked by a dominant negative PKC $\Delta$ l. *J Biol Chem*. 1996;271(49):31262–8.
  48. Bode AM, Ya W, Ma YJ, Surh Z. Inhibition of epidermal growth factor-induced cell transformation and activator protein 1 activation by [6]-gingerol. *Cancer Res*. 2001;61(3):850–3.
  49. Bode AM, Dong Z. Ginger. CRC Press; 2004. p. 157–68.
  50. Nigam N, Bhui K, Prasad S, George J, Shukla Y. Gingerol induces reactive oxygen species regulated mitochondrial cell death pathway in human epidermoid carcinoma A431 cells. *Chem Biol Interact*. 2009;181(1):77–84.

51. Nigam N, George J, Shukla Y. Molecular Targets and Therapeutic Uses of Spices. In *molecular targets and therapeutic uses of spices: Modern Uses for Ancient Medicine*. 2009;p. 225–6.
52. Keum Y, Sam J, Kim K, Lee KH, Park YJ, Surh JN. Induction of apoptosis and caspase-3 activation by chemopreventive [6]-paradol and structurally related compounds in KB cells. *Cancer lett*. 2002;177(1):41–7.
53. Wei QY, Ma JP, Cai YJ, Yang L, Liu ZL. Cytotoxic and apoptotic activities of diarylheptanoids and gingerol-related compounds from the rhizome of Chinese ginger. *J Ethnopharmacol*. 2005;102(2):177–84.
54. Lee H, Young SH, Park M, Lee HJ, Kim Y, Shi Y. 1-Dehydro-[10]-gingerdione from ginger inhibits IKK $\beta$  activity for NF- $\kappa$ B activation and suppresses NF- $\kappa$ B-regulated expression of inflammatory genes. *Brit J Pharmacol*. 2012;167(1):128–40.
55. Pan M, Hsiung MC, Hsieh JM, Kuo CS, Lai H, Wu S. 6-Shogaol induces apoptosis in human colorectal carcinoma cells via ROS production, caspase activation, and GADD 153 expression. *Mol Nutr Food Res*. 2008;52(5):527–37.
56. Miranda M, Sun X, Assis OBG, Ference C, Ferreira MD, Baldwin EA. Antifungal activity of Zingiber officinale Roscoe (ginger) oil and extracts, associated with carnauba wax nanoemulsions, on fungal control of harvest papaya. *Int Soc Horticultural Sci*. 2019;1325:191–8.
57. Li X, Li CC, Kristine CY, Mcgrath VH, Tran YM. Attenuation of proinflammatory responses by S-[6]-gingerol via inhibition of ROS/NF-kappa B/COX2 activation in HuH7 cells. *Evid Based Complementary Altern Med*. 2013;p. 146142. doi:10.1155/2013/146142.
58. Kim S, Joydeb OK, Kundu YK, Shin JH, Park MH, Cho TY. Gingerol inhibits COX-2 expression by blocking the activation of p38 MAP kinase and NF- $\kappa$ B in phorbol ester-stimulated mouse skin. *Oncogene*. 2005;24(15):2558–67.
59. Kim S, Joydeb OK, Kundu YK, Shin JH, Park MH, Cho TY. Gingerol inhibits COX-2 expression by blocking the activation of p38 MAP kinase and NF- $\kappa$ B in phorbol ester-stimulated mouse skin. *Oncogene*. 2005;24(15):2558–67.
60. Salariya AM, Habib F. Antioxidant activity of ginger extract in sunflower oil. *J S Food Agriculture*. 2003;83(7):624–9.
61. Takada Y, Murakami A, Aggarwal BB. Zerumbone abolishes NF- $\kappa$ B and I $\kappa$ B $\alpha$  kinase activation leading to suppression of antiapoptotic and metastatic gene expression, upregulation of apoptosis, and downregulation of invasion. *Oncogene*. 2005;24(46):6957–69.
62. Ichikawa H, Takada Y, Murakami A, Aggarwal BB. Identification of a novel blocker of I $\kappa$ B $\alpha$  kinase that enhances cellular apoptosis and inhibits cellular invasion through suppression of NF- $\kappa$ B-regulated gene products. *J Immunol*. 2005;174(11):7383–92.
63. Kim M, Seo J. Ginger attenuates inflammation in a mouse model of dextran sulfate sodium-induced colitis. *Food Sci Biotechnol*. 2018;27(5):1493–501.
64. Im JA, Kim MS, Kwon O, Kim JY. Animal model of intestinal anti-inflammatory effect of ginger-cinnamon complex. *Food Sci Biotechnol*. 2021;30(9):1249–56.
65. Li C, Li J, Jiang F, Tzvetkov NT, Horbanczuk JO, Li Y. Vasculoprotective effects of ginger (Zingiber officinale Roscoe) and underlying molecular mechanisms. *Food Function*. 2021;12(5):1897–913.
66. Srivastava KC, Mustafa T. Ginger (Zingiber officinale) in rheumatism and musculoskeletal disorders. *Med Hypotheses*. 1992;39(4):342–8.
67. Gomar A, Hosseini A, Mirazi N. Memory enhancement by administration of ginger (Zingiber officinale) extract on morphine-induced memory impairment in male rats. *J Acute Dis*. 2014;3(3):212–7.
68. Choi J, Jeong MS, Sun Y, Kim M. Pharmacotherapeutic potential of ginger and its compounds in age-related neurological disorders. *Pharmacol Therapeutics*. 2018;182:56–69.
69. Choi J, Jeong MS, Sun Y, Kim M. Pharmacotherapeutic potential of ginger and its compounds in age-related neurological disorders. *Pharmacol Ther*. 2018;182:56–69.
70. Dias MC, Spinardi-Barbisan ALT, Rodrigues M, De Camargo J, Teran E, Barbisan LF. Lack of chemopreventive effects of ginger on colon carcinogenesis induced by 1, 2-dimethylhydrazine in rats. *Food Chem Toxicol*. 2006;44(6):877–84.
71. Bidinotto L, Tadeu AT, Barbisan NS, Rocha DS, Fávero M, Salvadori LF. Effects of ginger (Zingiber officinale Roscoe) on DNA damage and development of urothelial tumors in a mouse bladder carcinogenesis model. *Environ Mol Mutagenesis*. 2006;47(8):624–30.
72. Bode AM, Dong Z. The amazing and mighty ginger. *Herb Med Biomol Clin Aspects*. 2011;47(8):624–30.

### Author biography

**Amit Gajanan Nerkar**, Professor and Research Head  
 <https://orcid.org/0000-0002-1377-8466>

**Srushti Ghadge**, Student

**Cite this article:** Nerkar AG, Ghadge S. Ethnopharmacological review of ginger for anticancer activity. *Curr Trends Pharm Pharm Chem* 2022;4(4):158-164.